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Abstract

Food adulteration is a major problem, which has been of concern for years to both consumers and regulators. In order to increase their profits, businesses use fraudulent practices, most often involving either the substitution of a high value ingredient for a lower value ingredient or the absence of ingredients which, according to the type of the product and its labelling, should be present in the product. Adulteration results in consumer deception and financial loss, leading to a lack of consumer confidence in certain producers. Furthermore, for certain groups of consumers, such fraud may lead to health problems.

In this paper, after giving the definition of adulteration, reference is made to the Greek and European legislation regarding food fraud and proper labelling. It then refers to some cases of adulteration which have been of public concern and many of which have contributed to the increased vigilance of the control authorities. Finally, the molecular methods most commonly used in the detection of food adulteration are analyzed and several studies in which these methods have been used in a range of food and feed categories are reported together with their results.

1.3.1

(Singuluri & Sukumaran, 2014). (Singh & Gandhi, 2015)

(Singuluri & Sukumaran, 2014). (Singh & Gandhi, 2015).

(Fischer et al., 2011).

(DDT), (HCB)

(Fischer et al., 2011).

(H₂O₂), (K₂Cr₂O₇), (HgCl₂).

(Singh & Gandhi, 2015).

- 1.
- 2.
- 3.

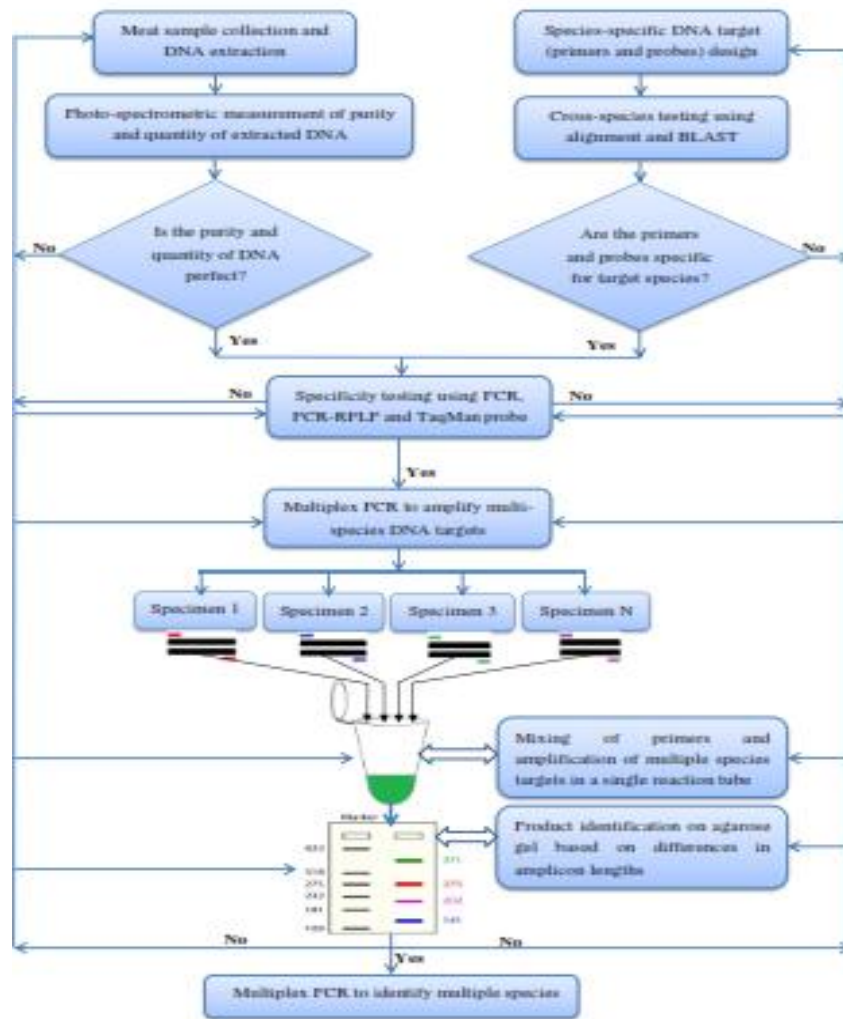
PCR
 Mullis Erlich Cetus
 1980 (John M.S. Bartlett & David Stirling, 2003).

DNA *Escherichia coli*. PCR
 DNA PCR, DNA
 E. coli, DNA
 95°C *Thermophilus aquaticus*
 Cetus Gelfand
 PCR 72°C, DNA
 E. coli, DNA PCR.
 Taq DNA Cetus Perkin-Elmer
 PCR (), PCR (John M.S. Bartlett & David Stirling, 2003).

2.3

μ (Multiplex PCR)

Chamberlain et al. (1988) μ μ
PCR μ
Duchenne. , Multiplex PCR ,
μ
μ (Eaqub et al., 2014).
μ (PCR) μ PCR
μ
μ
Multiplex PCR μ μ
(Markoulatos et al., 2002).
Multiplex PCR μ μ
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μ μ μ μ μ
Multiplex PCR
μ μ μ μ μ , PCR
, - μ μ , μ
μ
10⁷ μ μ .
μ , 0,5 μ
μ μ - μ μ ,
(Markoulatos et al., 2002).
, Multiplex PCR
μ μ μ μ μ .
μ , μ μ μ μ
PCR (μ control),
μ μ PCR. μ ,
μ μ Multiplex PCR
control μ μ
μ . Multiplex PCR,
μ μ μ (PCR PCR-RFLP.
silico in vitro (Markoulatos et al., 2002). μ μ in



Εικόνα 3: Σχηματική παρουσίαση των διαφόρων βημάτων σε μια Multiplex PCR. Στο διάγραμμα, το Yes υποδηλώνει ικανοποιητικά αποτελέσματα και No υποδηλώνει μη ικανοποιητικό αποτέλεσμα που χρειάζεται βελτιστοποίηση ή επανάληψη προηγούμενων βημάτων όπως φαίνονται με τα βέλη (Eaqub et al., 2014).

Multiplex PCR
Matsunaga et al. (1999)
PCR
b (*cytb*),
multiplex PCR
b (*cytb*), 12S RNA (rRNA), 16S rRNA, D-loop, transfer RNA (tRNA)-Val, ND5, ND2, ATPase6/ATPase8, 18S rRNA, (SINE), (LINE).

Multiplex PCR (multiplex end-point PCR) .

multiplex end-point PCR .

SYBR Green TaqMan probe.

Multiplex PCR (Eaqub et al., 2014).

2.4 DNA Barcoding

«Barcoding Life» 12-19 2005 (David E. Schindel, 2005).

50 «DNA Barcoding» 2003 (Paul Hebert, 2003).

(Savolainen et al., 2005).

DNA Barcoding DNA (Savolainen et al., 2005).

DNA Barcoding (Nicolas Hubert & Robert Hanner, 2015).

DNA, PCR, DNA Barcoding .

Barcoding et al., 2012). DNA barcode. ITS, DNA (Galimberti et al., 2012).

2.5 RAPD (Rapid Amplification of Polymorphic DNA)

RAPD ("rapid"), Random Amplification of Polymorphic DNA, PCR, DNA (N. Senthil Kumar & G. Gurusubramanian, 2011).

(10 μg) RAPD DNA PCR. (Nandani Kumari & Saroj Kumar Thakur, 2014). [10 μg DNA (DAF)] PCR (N. Senthil Kumar & G. Gurusubramanian, 2011).

DNA, DNA, DNA (Nandani Kumari & Saroj Kumar et al., 2014).

DNA";
 DNA.
 DNA (),
) (Nandani Kumari & Saroj Kumar Thakur, 2014).

2.6 (NGS)

DNA RNA.
 DNA Human Genome Project,
 Human Genome Project 13 2003.
 Sanger. Sanger (Edward Sanger,
 1975
 NGS (NGS).
 DNA
 NGS
 (Ayman Grada & Kate Weinbrecht, 2013).

NGS,
 DNA
 Illumina.
 DNA
 Illumina
 DNA (" "),
 Illumina iSeq 100,
 1,2
 HiSeq
 X Ten,
 1,8

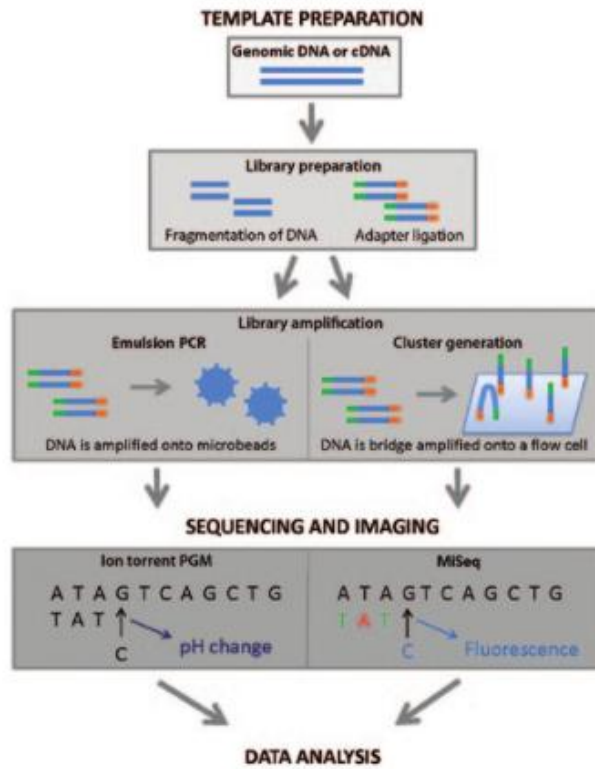
Illumina 2×300 MiSeq.
 Ion Torrent,
 Ion Torrent H⁺
 DNA.
 Machine (PGM), Proton, GeneStudio S5.

(SMRT) Pacific Biosciences (PacBio).
 DNA
 PacBio
 [60,000 (60, kb)].
 PacBio Sequel,
 DNA

Nanopore Technologies, MinION
 PromethION (SmidgION,
 smart phone
 DNA
 PCR (Haynes et al., 2019).

GenBank SRA (Sequence Read Archive).
 JRC GMO-Amplicons
 240,000 *Silico*
 barcode

NGS
 NGS



Εικόνα 4: μεθοδολογία αλληλούχησης επόμενης γενιάς (Ayman Grada & Kate Weinbrecht, 2013)

NGS

FDA,

NGS

NGS

NGS

(Haynes et al., 2019).

2.7

(HRM)

DNA.

(Tm)

Tm

2 °C,

PCR

GC. (Maria Erali & Carl Wittwer, 2010).

DNA PCR. (Maria Erali & Carl Wittwer, 2010).

DNA μ (ngs), PCR. μ
 LightCycler μ 1997. SYBR Green I μ
 μ (Maria Erali & Carl Wittwer, 2010).

HRM μ (Tm), PCR μ .
 μ (Carl T. Wittwer, 2009). HRM μ
 μ : μ SYBR Green I μ
 μ m, . . μ
 μ μ SYBR Green I.
 μ HRM μ
 μ , μ

de novo . LC-Green μ .
 μ μ μ LCGreen+ (Idaho Technology Inc.), Syto9 (Invitrogen, Carlsbad, CA), EvaGreen (Biotum) LightCycler 480 ResoLight Dye (Roche, Indianapolis, IN).

et.al, 2009). μ μ μ PCR (Vossen μ
 μ μ μ , μ μ μ
 . , μ μ μ μ μ
 μ μ μ μ μ μ μ
 (Maria Erali & Carl Wittwer, 2010).

μ μ μ HRM. μ , μ
 μ μ . μ μ μ μ
 μ . μ μ μ μ , Seipp et al. (2007)
 μ μ μ μ μ : μ μ μ
 μ μ μ μ μ μ μ
 Tm 38%, μ μ μ μ μ μ μ
 μ μ μ μ μ HRM .

Cervus canadensis ()
C. elaphus ().

DNA.

DNA Barcoding

,

(Galimberti et al., 2013).

DNA

DNA

Barcoding

DNA: i)

DNA (

ii)

DNA

(Aslan et al., 2009).

mtDNA

(Galimberti et al., 2013).

DNA Barcoding

DNA, 650

Sanger (bp).

DNA

200 bp.

Sanger

DNA Barcoding

Sanger

(NGS)

DNA Barcoding

DNA metabarcoding,

NGS

DNA
(Cottenet et al., 2020).

3.1.2

2019 Ran-Ran Xing et al., DNA metabarcoding
DNA metabarcoding.

Gallus gallus.
Sanger
(27)
16S RNA (16S rRNA).
(Sus scrofa)

μ μ μ (Salmo salar)
(Thunnus tonggol), μ μ ο
μ μ P3 (μ)
: (44,75%), (14,87%)
) (14,05%),
μ μ .
μ μ .
μ NGS Food Authenticity Workflow (Thermo Fisher
Scientific) μ μ μ μ
(Cottenet et al., 2020). μ μ μ μ
μ μ , μ μ μ μ
μ μ . , μ μ μ μ
NGS ,45 μ μ μ
(30) (15) μ μ μ
NGS, μ μ μ μ μ
μ μ μ μ μ
, , , , 2 μ .

Πίνακας 2: Καθαρά είδη κρέατος που ελέγχθηκαν με NGS.

Tested species	Reported by NGS	Tested species	Reported by NGS
Beef	<i>Bos taurus</i>	Chicken	<i>Gallus gallus</i>
Bison	<i>Bison bison</i>	Turkey	<i>Meleagris gallopavo</i>
Water buffalo	<i>Bubalus bubalis</i>	Guinea fowl	<i>Numida meleagris</i>
Yak	<i>Bos grunniens</i>	Muscovy duck	<i>Cairina moschata</i>
Pork	<i>Sus scrofa</i>	Mallard duck	<i>Anas platyrhynchos</i>
Horse	<i>Equus caballus</i>	Goose	<i>Alopochen aegyptiacus</i>
Donkey	<i>Equus asinus</i>	Pigeon	<i>Columba livia</i>
Zebra	<i>Equus quagga</i>	Pheasant	<i>Phasianus colchicus</i>
Sheep	<i>Ovis aries</i>	Partridge	<i>Alectoris chukar</i>
Goat	<i>Capra hircus</i>	Emu	<i>Dromaius novaehollandiae</i>
Hare	<i>Lepus capensis</i>	Ostrich	<i>Struthio camelus</i>
Rabbit	<i>Oryctolagus cuniculus</i>	Daw	<i>Corvus macrorhynchos</i>
Fallow deer	<i>Cervus dama</i>	Lama	<i>Lama glama</i>
Red deer	<i>Cervus elaphus</i>	Springbok	<i>Antidorcas marsupialis</i>
Reindeer	<i>Rangifer tarandus</i>	Impala	<i>Aepyceros melampus</i>
Roe deer	<i>Capreolus capreolus</i>	Kudu	<i>Tragelaphus strepsiceros</i>
Fox	<i>Vulpes vulpes</i>	Bushbuck	<i>Tragelaphus scriptus</i>
Weasel	<i>Mustela erminea</i>	Camel	<i>Camelus bactrianus</i>
Muskrat	<i>Ondatra zibethicus</i>	Red kangaroo	<i>Macropus rufus</i>
Badger	<i>Meles meles</i>	Common wallaroo	<i>Macropus robustus</i>
Dog	<i>Canis familiaris</i>	Oryx	<i>Oryx leucoryx</i>
Cat	<i>Felis catus</i>	Gnu	<i>Alcelaphus buselaphus</i>
Rat	<i>Rattus norvegicus</i>	Crocodile	<i>Crocodylus niloticus</i>

Πίνακας 3: Ανίχνευση και ταυτοποίηση ειδών κρέατος σε δείγματα εμπλουτισμένα με 1% (w/w).

Matrix	Reads	Adulterant	Reads
<i>Bos taurus</i>	24873	<i>Ovis aries</i>	163
<i>Bos taurus</i>	23659	<i>Capra hircus</i>	612
<i>Bos taurus</i>	24770	<i>Lepus capensis</i>	1897
<i>Bos taurus</i>	28501	<i>Oryctolagus cuniculus</i>	245
<i>Bos taurus</i>	26798	<i>Macropus rufus</i>	214
<i>Bos taurus</i>	27127	<i>Capreolus capreolus</i>	289
<i>Bos taurus</i>	27529	<i>Cervus elaphus</i>	759
<i>Bos taurus</i>	29764	<i>Rangifer tarandus</i>	573
<i>Bos taurus</i>	24108	<i>Antidorcas marsupialis</i>	379
<i>Bos taurus</i>	27369	<i>Equus hemionus</i>	286
<i>Bos taurus</i>	24250	<i>Lama glama</i>	209
<i>Bos taurus</i>	22528	<i>Gallus gallus</i>	325
<i>Bos taurus</i>	25992	<i>Canis familiaris</i>	974
<i>Bos taurus</i>	25803	<i>Bison bison</i>	223
<i>Bos taurus</i>	32428	<i>Bison bison</i>	312
<i>Bos taurus</i>	20959	<i>Cervus dama</i>	323
<i>Bos taurus</i>	106117	<i>Tragelaphus strepsiceros</i>	1433
<i>Bos taurus</i>	89773	<i>Felis catus</i>	717
<i>Bos taurus</i>	87641	<i>Rattus norvegicus</i>	701
<i>Bos taurus</i>	86795	<i>Vulpes vulpes</i>	1624
<i>Gallus gallus</i>	102451	<i>Meleagris gallopavo</i>	587
<i>Gallus gallus</i>	114141	<i>Catrina moschata</i>	1953
<i>Gallus gallus</i>	101648	<i>Alopochen aegyptiacus</i>	1661
<i>Gallus gallus</i>	113967	<i>Struthio camelus</i>	1109
<i>Gallus gallus</i>	122546	<i>Oryctolagus cuniculus</i>	2453
<i>Gallus gallus</i>	110329	<i>Columba livia</i>	3963
<i>Gallus gallus</i>	130693	<i>Numida meleagris</i>	899
<i>Bos taurus</i>	27818	<i>Bubalus bubalis</i>	176
		<i>Equus caballus</i>	157
		<i>Sus scrofa</i>	196
<i>Ovis aries</i>	28313	<i>Capra hircus</i>	297
		<i>Sus scrofa</i>	276
Cooked sample (n = 20)			
<i>Bos taurus</i>	Detected 20/20	<i>Rangifer tarandus</i>	Detected 20/20
		<i>Sus scrofa</i>	Detected 19/20

μ μ
 1% (w/w),
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 , μ 4
 NGS
 μ μ
 , μ μ μ μ μ μ
 1% . μ 4
 μ μ

μ , μ 12S rDNA
 PCR μ 410 bp.
 BLASTn μ
 12S rDNA μ 99-100%
 μ *Bos taurus*
 , μ 12S rDNA μ
 μ . μ >98%
 GenBank. μ ,
 12S rDNA. , μ Multiplex PCR μ
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 μ μ halal .
 μ (100%), μ
 " " , 50 80 (62,5%)
 μμ . , μ
 μ μ 61,6% μ
 μ 100% μ
 . , 62,5% μ
 μ ,

3.1.4 μ HRM

μ μ , μ μ μ
 μ . μ , μ (3.1.3) μ
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 "Halal" μ μ μ μ μ
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 μ μ μ μ μ μ
 μ μ μ , μ μ 1,9 μμ
 μ . μ μ μ μ

FISH-BOL (Barcode Of Life Data).

DNA Barcoding

DNA barcoding is a method of species identification using short DNA sequences. It is widely used in taxonomy and systematics. The process involves comparing a DNA sequence from an unknown specimen to a reference database. This method is particularly useful for identifying species that are difficult to distinguish based on morphology. The reference database is often built from known species, and the DNA sequence of the specimen is compared to these sequences to determine its identity. This process is often automated using software tools. The resulting DNA barcode can be used to identify the species and to study its evolutionary relationships. This method is also used in forensic science and in the study of invasive species. The DNA barcode is a unique identifier for each species, and it can be used to track the movement of species across different regions. This method is also used in the study of biodiversity and in the conservation of endangered species. The DNA barcode is a powerful tool for species identification and for understanding the relationships between different species. It is a key component of modern taxonomy and systematics. The DNA barcode is a unique identifier for each species, and it can be used to track the movement of species across different regions. This method is also used in the study of biodiversity and in the conservation of endangered species. The DNA barcode is a powerful tool for species identification and for understanding the relationships between different species. It is a key component of modern taxonomy and systematics.

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(*Oreochromis spp.*), (*Lates niloticus*) (*Pangasius Pangasionodon spp.*) (: Cichlidae, : Perciformes)

(: Latidae, : Perciformes),

(: Pangasiidae, : Siluriformes)

DNA Barcoding

μ μ μ μ

(L. niloticus), (Pangasius

Pangasionodon spp.) (Oreochromis spp.) , μ :

μ μ 2013, :

, Monufia Qalyubia. μ μ

7. μ μ .

Πίνακας 7: Αριθμός δειγμάτων που ελήφθησαν για κάθε είδος, πληροφορίες που αναγράφονται στην ετικέτα και τοποθεσίες συλλογής.

Number of samples	On label		Origin of product	Sampling location (province)
	Common name (expected species)	Processed fish product		
20	Nile perch (<i>Lates niloticus</i>)	Frozen Fillets	Egypt	Monufia and Qalyubia, Egypt
20	Tra fish (<i>Pangasius hypophthalmus</i>)	Frozen Fillets	Vietnam	Cairo, Egypt
20	Basa (<i>Pangasius bocourti</i>)	Frozen Fillets	Vietnam	Monufia, Egypt
30	Tilapia (<i>Oreochromis spp.</i>)	Frozen Fillets	Egypt	Monufia, Cairo and Qalyubia, Egypt

μ (77%) μ μ

μ μ (

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μ (GenBank BOLD), μ (5).

μ μ 50%

μ μ 33,3% μ μ ,

tra () (P.

hypophthalmus). ο , Basa (*P. bocourti*)

100% tra (*P. hypophthalmus*), , μ

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 2017. *Sciaenops ocellatus* μ 90% μ
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Ocellatus. μ , μ μ μ μ
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alata, μ μ (*Upeneus japonicus, Lepidotrigla japonica, L. Saurida Trachurus*).
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Xue Yu μ μ μ μ μ
Xue Yu, μ μ μ μ μ μ μ
 , *Xue Yu,* μ μ μ
Xue Yu, μ μ μ
 Gadidae
 Gadiformes. μ μ μ μ μ μ μ
 Gadiformes *Xue Yu.*
 μ μ μ μ μ μ μ μ
 μ , *Xue Yu,* μ μ μ μ μ 153
 μ 30 μ μ μ , 16 . O
 μ DNA Barcoding (FDB MDB)
 (Xiong et al., 2018).

Gadidae 3) *Xue Yu* Gadiformes 2) *Xue Yu*
Gadus spp. *Xue Yu*,
 58% (1 2) 100% (3).
Lagocephalus spp., pufferfish,
 37 (TTX), pufferfish
 pufferfish.
 pufferfish
 pufferfish
 pufferfish
 1990,
 pufferfish.
 pufferfish, 2010,
 pufferfish
 2016. *T. rubripes*,
obscurus *Takifugu*
Lagocephalus spp.
Lagocephalus spp. *Xue Yu*
 Ommastrephidae Loliginidae Myopsina.
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3.2.2

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(NGS)

(Holothuroidea)

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Apostichopus japonicus.

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Xing et al. (2021)

μ

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NGS

μ μ DNA mini-barcoding

μ

40

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23

mini-barcode.

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12

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 . μ H. Tubulosa H.
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 Apostichopus japonicus,
 Cucumaria frondosa. μ Apostichopus japonicus
 Cucumaria frondosa, μ μ .
 μ NGS
 μ . μ (μ
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 A. japonicus, . μ ,
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 2016 μ μ μ μ
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 (Carvalho et al., 2017).
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 , μ μ μ μ
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 μ μ μ , μ μ μ μ μ μ
 μ μ μ "μ " μ :
 Gadus macrocephalus, Gadus morhua, Gadusogac, Boreogadus saida.
 Gadus morhua μ (. .
 Gadus macrocephalus),
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 μ μ μ DNA Barcoding.
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 μ μ (rtPCR) μ
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 μ μ DNA (NGS), μ
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 Belo Horizonte (μ). μ
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 S.I.F. μ μ
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 (Instrução Normativa
 No 29, 2015) FishBase (www.fishbase.org) μ
 μ μ μ μ μ μ (59%)
 μ μ μ μ μ μ (18%)
 μ μ μ μ μ μ (23%) μ
 . μ μ μ μ μ μ 41% (N=9)
 μ μ μ μ μ μ S.I.F.
 μ μ 22
 . μ μ (*Gadus morhua* *Gadus*

μ μ / DNA .
 μ μ BDR CR, . . μ
 μ , μ
 μ μ , μ μ
 μ μ μ .
 μ , μ .

3.2.3 μ μ PCR

38% μ 2015. , μ
 μ , μ
 μ μ . , μ
 3.2.1, μ μ , μ
 μ μ μ μ μ μ
 μ , μ μ μ Gadiformes
 (μ μ μ *G. morhua*, *G. macrocephalus* *G. chalcogrammus*)
 μ μ Gadiformes
 μ Gadiformes
Pollachius virens, *Melanogrammus aeglefinus*, *Merluccius merluccius*, *Merluccius australis* *Albatrossia pectoralis*.

, Rongzhen Shi et al. (2020) μ μ μ
 mini DNA barcoding HRM μ
Gadus Gadiformes. μ mini-barcoding
 μ 12S rRNA , μ
 μ μ HRM. μ ,
 μ μ μ μ ,
 μ μ μ .
 μ μ μ μ
 μ Gadiformes (*G. morhua*, *G. macrocephalus*, *G. chalcogrammus*, *Pollachius virens*, *Melanogrammus aeglefinus*, *Merluccius merluccius*, *Merluccius australis* *Albatrossia pectoralis*) 33 μ Gadiformes
 μ , μ . ,

μ DNA Barcoding. Nanjing
 μ , 26 μ
 (9). μ

Πίνακας 9: Εφαρμογή της αναπτυχθείσας μεθόδου σε εμπορικά δείγματα (οι σκούρες λέξεις είναι τα δείγματα που ταυτοποιήθηκαν ως είδη έξω από την τάξη των Gadiformes)

Sample code	Real time PCR			DNA barcoding analysis	
	Cq±SD	HRM cluster	Confidence (%)	NCBI (similarity value ≥ 98%)	BLOD (similarity value ≥ 98%)
S1	21.20±0.38	<i>Gadus</i>	99.8±0.2	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
S2	20.58±0.07	<i>Gadus</i>	99.0±1.2	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
S3	21.48±0.37	<i>Gadus</i>	99.2±0.7	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
S4	34.14±0.58	NRC	–	<i>R. ectenurus</i>	<i>R. ectenurus</i>
S5	19.25±0.07	<i>Gadus</i>	99.5±0.6	<i>G. morhua</i>	<i>G. morhua</i>
S6	19.50±0.05	<i>Gadus</i>	99.5±0.5	<i>G. morhua</i>	<i>G. morhua</i>
S7	33.82±1.87	NRC	–	<i>L. litulon</i>	<i>L. litulon</i>
S8	33.05±0.38	NRC	–	<i>U. japonicus</i>	<i>U. japonicus</i>
S9	19.45±0.11	<i>Gadus</i>	99.6±0.0	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
S10	18.87±0.37	<i>Gadus</i>	98.8±0.0	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
S11	17.09±0.28	<i>Gadus</i>	97.8±0.5	<i>G. morhua</i>	<i>G. morhua</i>
S12	19.83±0.60	<i>Gadus</i>	98.7±0.2	<i>G. morhua</i>	<i>G. morhua</i>
YZ2	33.20±0.18	NRC	–	<i>Lagocephalus</i>	<i>Lagocephalus</i>
YZ3	19.38±0.17	<i>Gadus</i>	99.0±0.4	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
YZ6	18.48±0.57	<i>Gadus</i>	99.8±0.0	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
YZ7	19.14±0.68	<i>Gadus</i>	99.2±0.6	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
YZ9	20.25±0.22	<i>Gadus</i>	99.6±0.2	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
YZ11	20.50±0.15	<i>Gadus</i>	99.7±0.1	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
YZ13	34.32±1.07	NRC	–	<i>L. litulon</i>	<i>L. litulon</i>
YZ15	33.05±0.18	NRC	–	<i>L. litulon</i>	<i>L. litulon</i>
YZ17	19.45±0.51	<i>Gadus</i>	99.0±0.6	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
YZ18	20.47±0.37	<i>Gadus</i>	98.6±0.9	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
YZ19	21.43±0.18	<i>Gadus</i>	99.6±0.3	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
YZ20	18.83±0.20	<i>Gadus</i>	98.6±0.9	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
YZ27	19.23±0.41	<i>Gadus</i>	98.8±0.6	<i>T. chalcogramma</i>	<i>T. chalcogramma</i>
YZ28	33.53±0.77	NRC	–	<i>Lagocephalus</i>	<i>Lagocephalus</i>

NRC no reference cluster

μ *Gadus*, μ , μ μ μ
 μ S4, S7, S8, YZ2, YZ13, YZ15 YZ28 (9). μ μ
 HRM μ μ
 μ μ , DNA
 mini-barcoding, μ μ *Gadus* G.
chalcogrammus *G. morhua*, μ μ
 Rhynchoconger *ectenurus* (Anguilliformes, Congridae), *Lophius*

Yu-Min Lee et al. (2022) Multiplex PCR

14 μ (Octopus cyanea), (Enteroctopus dofleini) (Octopus vulgaris), 14 μ (Octopus minor), webfoot (Octopus ocellatus), (Sepia esculenta), (Todarodes pacificus), (Illex argentinus), jumbo (Dosidicus gigas), diamondback (Thysanoteuthis rhombus), swordtip (Loligo edulis), spear (Loligo bleekeri), (Loliolus japonica), beka (Loliolus beka) (Sepioteuthis lessoniana).

Multiplex PCR

30 μ parboiled, sushi, stir-fried, salted, takoyaki, (10). (10). PCR

PCR

Πίνακας 10: Εφαρμογή και επικύρωση της μεθόδου Multiplex PCR σε εμπορικά προϊόντα.

No	Product type	Origin	Multiplex PCR results		
			Big blue octopus	Giant Pacific octopus	Common octopus
1	Parboiled	Korea			+ + +
2	Parboiled	Korea		+ + +	
3	Parboiled	Korea			+ + +
4	Parboiled	Korea			+ + +
5	Parboiled	China			+ + +
6	Sushi	Philippine	+ + +		
7	Sushi	Vietnam	+ + +		
8	Salted	Korea	+ + +		
9	Stir-fried	Mauritania			+ + +
10	Salted	Korea			+ + +
11	Fish cake	NL		+ + +	
12	Dumpling	NL		+ + +	
13	Dumpling	Philippine	+ + +		
14	Stir-fried	Philippine	+ + +		
15	Stir-fried	Mauritania			+ + +
16	Nugget	Philippine	+ + +		
17	Salad	Korea			+ + +
18	Boiled	Mauritania			+ + +
19	Dumpling	China	+ + +		
20	Dried	Korea			+ + +
21	Parboiled	Korea			+ + +
22	Sushi	NL			+ + +
23	Fish cake	China	+ + +		
24	Takoyaki	China			+ + +
25	Porridge	Indonesia	+ + +		
26	Porridge	China	+ + +		
27	Dried	Korea		+ + +	
28	Roasted	Korea		+ + +	
29	Fried	Korea		+ + +	
30	Boiled	Korea		+ + +	

The test was independently three times using three different instruments.

NL, no label

+ means a positive result

3.3

μ

μ

μ

μ

μ

μ

μ

μ

μ

μ

μ

μ

μ

μ

μ

(Galimberti et al., 2013).

barcode, DNA barcode, INDEL barcodes (/) , Ayse Ozgur Uncu Ali Tevfik Uncu (2020) DNA Barcoding INDEL barcode PCR-CE (PCR-) (ghee) (1,5%) (*Zea mays* L.), (*Glycine max* (L.) Merr.), (*Brassica napus* L.) (*Helianthus annuus* L.) Bahri Dagdas International Agricultural Dagdas (Konya, (refined) (UAA) barcode *trnL* barcode PCR, P6 PCR-CE P6 5% P6 2% Nested PCR-CE

) μ μ , (2:1:1
)
 μ DNA μ μ ,
 DNA μ (0,2% μ
 μ mtDNA) μ μ μ
 μ μ DNA crescita,
 ()
 μ .
 μ μ , DNA
 μ . DNA μ
 μ , μ
 μ , μ
 μ μ μ .
 μ μ μ .
 μ μ μ
 (0,6%-1,8%)
 mtDNA. μ DNA,
 μ μ μ
 μ μ . μ μ
 μ μ μ .
 μ μ mtDNA μ
 μ μ μ μ μ μ
 μ μ DNA μ
 μ μ , μ μ NGS μ
 DNA μ μ μ .

μ Fabaceae (*V. faba*) *major* (μ)
minor () μ , μ
μ . ” ” . , *L.*
clymenum μ μ *Lathyrus*, *Lathyrus cicera*, *Lathyrus*
ochrus *Lathyrus sativus*, μ
, - μ (ODAP)
μ μ .
μ μ , μ
13. μ
μ μ (*V. faba*) μ μ μ *Lathyrus*
(*Lathyrus cicera*, *L. ochrus* *L. sativum*)
” ” *Pisum*.
μ ” 10 μ : ”
” ” ” ” (13). μ

Πίνακας 13: Εμπορικά προϊόντα "Φάβα", *Lathyrus* και συναφή είδη που χρησιμοποιήθηκαν στην παρούσα μελέτη.

No. genotype	Species/type	Abbreviation	Source
1	Commercial 1 ("Fava")	Com1	Market
2	Commercial 2 ("Fava")	Com2	Market
3	Commercial 3 ("Fava")	Com3	Market
4	Commercial 4 ("Fava")	Com4	Market
5	Commercial 5 ("Fava")	Com5	Market
6	Commercial 6 ("Fava")	Com6	Market
7	Commercial 7 ("Fava Santorinis")	Com7	Local producers
8	Commercial 8 (putative "Fava Santorinis")	Com8	Market
9	Commercial 9 (putative "Fava Santorinis")	Com9	Market
10	Commercial 10 (putative "Fava Santorinis")	Com10	Market
11	<i>Lathyrus sativus</i>	Lsa	NAGREF
12	<i>Lathyrus clymenum</i>	Lcl	AUTH, Seed Bank
13	<i>Pisum sativum</i> subsp. <i>eliatum</i>	Psel	NAGREF
14	<i>Vicia faba</i> subsp. <i>minor</i>	Vfmin	NAGREF
15	<i>Vicia faba</i> subsp. <i>major</i>	Vfmaj	NAGREF
16	<i>Lathyrus cicera</i>	Lci	NAGREF
17	<i>Pisum sativum</i>	Psa	NAGREF
18	<i>Lathyrus ochrus</i>	Loc	NAGREF

μ μ (Bar-HRM) μ
μ *L.clymenum* ,
μ 1:100 μ μ
. , 75% μ , μ
μ μ , μ
" " (Com8, Com9 Com10).
μ [*Curcuma longa* L. (Zingiberaceae)],
μ μ ,
μ μ μ , ,
, μ , ,

DNA, DNA
 DNA (, , , .)
 PCR.
 40 50 μ μ μ
 10 mg/kg μ ,
 7 μ 10 50 50
 mg/kg μ . μ
 μ , μ , μ
 μ .
 μ μ
 μ premium
 , μ ,
 μ . , μ μ μ
 , μ μ , ,
 , μ μ μ .
 Vietina et al. (2013) μ μ μ DNA
 μ PCR μ
 HRM μ PCR. (*Olea europaea*
 L.), (*Corylus avellana* L.), (*Helianthus annuus* L.),
 (*Zea mays* L.), μ (*Sesamum indicum* L.) (*Cucurbita*
pepo L.), (*Glycine max* L.), (*Arachis hypogaea* L.) (*Oryza*
sativa L.). DNA μ
 , μ .
 μ Leccino μ μ "Il Posticcio"
 (Massa Carrara,). (, ,
) μ DNA
 μ PCR μ . DNA
 μ , μ
 μ .
 HRM , μ
 . HRM
 μ μ , μ μ Leccino, μ
 μ 10%.

Multiplex PCR. Dalmasso et al. (2004)

: () , () 4%
 4%, () , ()
 4%. 4%.

DNA. PCR Multiplex PCR

2014 Pegels et al.,
 PCR DNA (*Equus caballus*) TaqMan

(Facultad de Veterinaria, Universidad Complutense de Madrid,). 171
 (81 90)
 (μ μ μ),
 81 , 30 μ
 , 51 μ μ μ
 , DNA 30 μ

-) : / www.efet.gr
-) , . 4177 (8 2013)
-) , . 4619 (11 2019)
-) , . 71 (24 2018)
-) , . 4235 (11 2014)
-) () . 1830/2003 www.europa.eu
-) () . 1536/92 www.europa.eu
-) () . 1169/2011 www.europa.eu
-) () . 1379/2013 www.europa.eu
-) (EE) 625/2017 www.europa.eu
-) () . 1760/2000 www.europa.eu
-) () . 2065/2001 www.europa.eu
-) () . 178/2002 www.europa.eu
-) 2000/13/ www.europa.eu
-) () . 1379/2013 www.europa.eu
-) CODEX ALIMENTARIUS
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